Phylogenetic Differentiation of the Osprey *Pandion haliaetus* inferred from nucleotide sequences of the mitochondrial cytochrome b gene

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ABSTRACT

The mitochondrial cytochrome b gene of three of the four subspecies of the Osprey (*P. h. haliaetus*, *P. h. carolinensis*, and *P. h. cristatus*) was amplified by PCR and sequenced. Ospreys cluster as a monophyletic group within the Falconiformes and show a clear and unequivocal phylogeographic pattern according to the recognised subspecies. Genetic distances between the subspecies are between 1.9 and 3.8%, i.e. in the range of "good" species. Together with morphological characters and distribution pattern the genetic data indicate that the three osprey subspecies appear to represent distinct species.

INTRODUCTION

The Osprey *Pandion haliaetus* is a unique raptor that feeds entirely on fish and has a world-wide distribution. It has been placed in a monotypic family, the Pandionidae, within the Falconiformes (Sibley & Monroe 1990) and has been subdivided at present into four geographically defined subspecies:

- nominate *P. h. haliaetus* (Palaearctic, incl. Cape Verde Islands; migratory),
- P. h. carolinensis (North America; migratory),
- *P. h. ridgwayi* (Caribbean subregion; non-migratory),
- *P. h. cristatus* (Australia, from Sulawesi and Java to New Caledonia; nonmigratory)

When Ospreys were first described between 1822 (Vieillot 1822) and 1837 (Gould 1837), three species were recognised that were placed in the genus *Pandion*. Lumping started in 1874 (Sharpe 1874) and ended with Knowlton & Ridgway (1909) (summary Prevost 1983). In the following decades disputes mainly concerned the recognition of additional subspecies.

Ospreys vary in size, both geographically and by sex. Usually females are larger than males, about 20% larger in body mass, and 5-10% larger in wing length. Some overlap is possible, but this is limited; in North America, for example, nearly all males weigh less than 1600 grams, while nearly all females weigh more. Yet at a distance such differences are obscured and it is often difficult to distinguish between the sexes in the field, even though females tend to have darker breast plumage.

The overall habits and behaviour of the four subspecies are very similar. although a few characters have been described that differ (del Hoyo et al. 1994). The subspecies show only slight differences, mostly in plumage and size, with haliaetus and carolinensis the largest and darkest, and ridgwavi essentially a pale form of carolinensis. The Australian cristatus is a small osprey with dark breast band and a relatively pale crown (del Hoyo et al. 1994).

Table 1 summarises a few plumage characters and Table 2 differences in wing length as an indicator of size (Weick 1980; Prevost 1983; del Hoyo et al. 1994). The ospreys of the northern hemisphere appear to share a number of characters and differ from the Australasian subspecies P. h. cristatus, which is the smallest of the four subspecies.

Table 1. Morphological characters of the Pandion taxa

Sample sizes are in brackets (after Prevost 1983).

		Male	I	Female		
	Breast ¹	Crown ²	Breast	Crown		
	Average	average	average	average		
Palaearctic	2.7 (72)	2.5 (32)	3.4 (65)	2.7 (41)		
Cape Verde	2.2 (6)	2.5 (6)	2.3 (4)	2.5 (4)		
Red Sea	1.6 (5)	2.0 (5)	1.7 (3)	1.7 (3)		
North America	1.4 (49)	2.6 (49)	2.4 (47)	2.8 (46)		
Bahaman	1.0 (4)	1.0 (3)	1.0 (3)	1.0 (3)		
Australasian	2.9 (42)	1.5 (42)	3.7 (50)	1.3 (50)		

1 The breast markings were scored from 1 to 4: 1=the breast almost white; 2=a band of partially brown feathers < 3 cm wide; 3=a band of partially brown feathers > 3 cm wide; 4=a wide band of entirely brown feathers

2 Crown markings were scored from 1 to 3 : 1=less than 10 % black feathers; 2= from 10 to 50% black feathers; 3=over 50% black feathers

Another character is larger underwing coverts. All Palaearctic adult ospreys, and only these, have the large underwing coverts (one half rufous-brown, one half white). Although North American ospreys have underwings similar to those of Palaearctic ospreys, their larger underwing coverts are barred dark brown. In Bahamian ospreys these coverts are nearly white, whereas in Australasian birds they are nearly all ashy brown (Prevost 1983).

In this communication we report on a molecular phylogeny and phylogeography of ospreys based on nucleotide sequences of the mitochondrial b gene covering three of the subspecies. Experimental evidence is presented that the three taxa represent genetic lineages isolated for probably more than 1 million years.

Table 2. Wing lengths (init) of the Funation taxa (after Welck 1900)				
	females	males		
P. h. haliaetus	470-510	450-510		
P. h. carolinensis	488-518	462-506		
P. h. ridgwavi	455-495	433-483		
P. h. cristatus	425-490	426-431		

Table 2. Wing lengths (mm) of the Pandion taxa (after Weick 1980)

MATERIAL AND METHODS

We have isolated total DNA from feather, blood or tissue samples (see Wink 2000) which had been kindly supplied by several colleagues (B. Bed'Hom, J. Haapala, D. Ristow, J. Penhallurick, J.-C. Thibault, W. E. Boles, R. Schodde, O. Hadzofe). The cytochrome b gene was amplified by PCR (primer sequences in Wink & Sauer-Gürth 2000) and sequenced by using AlfExpress (Amersham Pharmacia Biotech) or ABI 3100 (Applied Biosystems) instruments. Sequences were aligned manually and analysed with the software packages PAUP* (Swofford, 2002) and MEGA2 (Kumar *et al.* 2001) (see Wink 2000; Wink & Sauer-Gürth 2000; Wink *et al.* 2002; Broders *et al.* 2003 for further details).

RESULTS AND DISCUSSION

Ospreys form a well-supported monophyletic clade (Fig. 1) (Wink & Sauer-Gürth 2000) and cluster within the basal groups of the Accipitridae; in other reconstructions *Pandion* takes a basal place outside the Accipitridae (which would support its status as a monotypic family). Several taxa of kites that had not been included into previous phylogenetic studies (Wink 2000; Wink & Sauer-Gürth 2000) take an even more basal position (Fig. 1) than hitherto seen for *Pandion*. However, longer mitochondrial sequences and nuclear sequences are needed to identify the position of *Pandion* unequivocally.

Table 3. Informative characters in the Pandion data set

	1111111 11222223344445556666667778888999990000000 992424499292468239128995694699144890012457
Taxon/Node	369756917799759274845049223448829306810831
P.h. haliaetus FL P.h. haliaetus Is P.b. haliaetus Co	GGGAATCCCGCGTCGATCGTGTGCACCACTTCGCTTAGTACC
P.h. haliaetus Po P.h. haliaetus Po P.h. haliaetus F20425	A
P.h. haliaetus F20429	
P.h. haliaetus F9263	
P.h. carolinensis USA 4252 P.h. carolinensis USA 4253	AGCATTAAACGAG.TAC.TGTCC.A.GTA AAGCATTAAACGAG.TA.ACATTA.CC.A.CTA
P.h. carolinensis USA 4255 P.h. carolinensis USA 4254	AAGCATTAAACGAG.TAC.TGTAACC.A.GTA AGCATTAAACGAG.TA.ACATTAACC.A.GTA
P.h. cristatus AUS 7011 P.h. cristatus AUS 21009	AAGTAC.AGCTAC.CG.TCCTCC
P.h. cristatus AUS 21009	.AAGTAC.AGCTAC.CG.TCCTCCG

Within the ospreys a clearcut phylogeographic pattern was found, irrespective of the methods or outgroups used for phylogeny reconstructions (MP, ML, NJ) (Fig. 2). The Australasian *P. h. cristatus* usually takes a basal position, whereas *P. h. carolinensis* and *P. h. haliaetus* cluster as sister taxa in MP reconstructions (this would agree with morphological characters). No phylogeographic pattern was found between different European populations of the osprey. Table 3 illustrates the comparably large number of phylogenetically informative characters in the *Pandion* data set.





Table 4 summarises the genetic distances between the three osprey taxa. Pdistances are between 2 and 4%; assuming a molecular clock of 2% = 1 million years of divergence (Tarr *et al.*, 1993; Wilson *et al.* 1987), the taxa have diverged 1 to 2 million years ago.

Table 4. Genetic distances between Pandion taxa				
	1*	2	3**	
1. P. h. haliaetus	-	2.1-3.5	1.9-2.6	
2, P. h. carolinensis	-	2.9-3.8		
3. P. h.cristatus	-			

*distances within European ospreys: 0.1-0.5%; **within Australian birds: 0.3-0.8%

Figure 2. Phylogenetic differentiation within Pandion haliaetus

MP= Maximum parsimony; NJ= Neighbour Joining (Kimura2-distance algorithm) Numbers at branches are bootstrap values in % (from 1000 replications)



Genetic distances between closely related sister species of some eagles (A. clanga/A. pomarina; A. heliaca/A. adalberti; H. fasciatus/H. spilogaster) are in the range of 1.7 to 2.1% (Table 5). Since the distances within ospreys clearly exceed this threshold, we suggest that the Pandion taxa represent "good" species although the morphological differences are small. They are also very small in case of the sister species mentioned in Table 5. Thus the old concept of the early 19th century (Vieillot 1822; Gould 1837), of defining at least three distinct osprey species, might not have been so wrong at all.

Table 5. Distance between closely related sister pairs of eagles (in %sequence divergence)

	1	2	3	4	5	6
1 Aquila clanga	-	1.7	7.1	7.5	7.2	6.7
2 Aquila pomarina		-	7.3	7.7	7.1	6.6
3 Aquila heliaca			-	1.8	7.4	6.6
4 Aquila adalberti				-	7.8	7.2
5 Hieraaetus fasciatus					-	2.1
6 Hieraaetus spilogaster						-

CONCLUSIONS

Genetic distances between osprey taxa are in a range that is typical for distinct raptor species. Morphological and genetic differences exist and support the suggestion, that the geographically defined subspecies may be recognised as distinct species:

- P. haliaetus
- P. carolinensis
- P. ridgwayi
- P. cristatus

REFERENCES

BRODERS, O., T. OSBORNE & M. WINK 2003. A mtDNA phylogeny of bustards (family Otitidae)based on nucleotide sequences of the cytochrome b gene. *J. Ornithol.* 144, 176-185

CRAMP, S. 1980. The Birds of the Western Palearctic. Vol. II. Oxford University Press.

DEL HOYO, J., A. ELLIOTT & J. SARGATAL 1994. Handbook of the Birds of the World. Vol. 2, Lynx Edicions, Barvelona

GOULD, J. 1837. Characters of a large number of new species of Australasian birds. Proc. Zool. Soc. 5, 138-157.

KNOWLTON, F.H. & R. RIDGWAY 1909. Birds of the World: A popular account. Frank Holt, New York.

KUMAR, S., K. TAMURA, I. B. JAKOBSEN & M. NEI 2001. MEGA2: Molecular Evolutionary Genetics Analysis software, Arizona State University, Tempe, Arizona, USA.

PREVOST, Y.A. 1983. Osprey distribution and subspecies taxonomy. In D.M. Bird (Ed.) Biology and Management of Bald Eagles and Ospreys: 157-174. Montreal: Raptor Research Foundation.

SEIBOLD, I., A. HELBIG, B.-U. MEYBURG, J. NEGRO & M. WINK 1996. Genetic differentiation and molecular phylogeny of European Aquila eagles according to cytochrome b nucleotide sequences. pp. 1-15, In Eagle studies (B.-U. Meyburg & R. Chancellor (Eds.). WWGBP, Berlin, London & Paris

SHARPE, R.B. 1874. Catalogue of birds, Vol 1: Catalogue of the Accipitres or Birds of Prey. BMNH Trustees, London.

SIBLEY, G.C. & J.E. AHLQUIST 1990. Phylogeny and Classification of Birds. A study in molecular evolution. Yale University Press, New Haven.

SIBLEY, C.G. & B.L. MONROE 1990. Distribution and Taxonomy of Birds of the World. Yale University Press, New Haven, 1990.

SWOFFORD, D.L. 2002. PAUP-Phylogenetic analysis using parsimony. Version PAUP*4.0b10.

TARR, C.L. & R.C. FLEISCHER 1993. Mitochondrial DNA variation and evolutionary relationships in the amakihi complex. Auk 110, 825-831

VIEILLOT, L.P. 1822. Galerie des oiseaux I: 33

WEICK, F. 1980. Die Greifvögel der Welt. P. Paray, Hamburg.

WILSON, A.C., H. OCHMAN & E.M. PRAGER 1987. Molecular time scale for evolution. Trends Genetics 3, 241-247, 1987

WINK, M. 2000. Advances in DNA studies of diurnal and nocturnal raptors. In *Raptors at Risk* (R.D. Chancellor & B.-U. Meyburg, Eds.) WWGBP/Hancock House. pp 831-844.

WINK, M. & H SAUER-GÜRTH 2000. Advances in the molecular systematics of African raptors In *Raptors at Risk* (R.D. Chancellor & B.-U. Meyburg, Eds.) WWGBP/Hancock House. pp 135-147

WINK, M., H. SAUER-GÜRTH & E. GWINNER 2000. A molecular phylogeny of stonechats and related turdids inferred from mitochondrial DNA sequences and genomic fingerprinting by ISSR-PCR. *British Birds* 95, 349-355

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